Title	RNA Extraction From mammalian cells		
Protocol Number	MOLBIO20001	Written by	Duha A.
Adapted from	MO Lab and thermo fisher	Version	1.0
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Make sure the centrifuge is at 4°C before you start the extraction

Reagents:

- TRIzol
- Chloroform
- Isopropanol
- 75% ETOH
- RNAase free water

Protocol:

- 1. To the eppendorfs containing TRIzol lysed cells, add 200μL of chloroform per 1mL of TRIzol
- 2. Mix by inverting the tube several times
- 3. Incubate for 2-3 mints at RT
- 4. Centrifuge the content of each eppendorf at 10,800 RPM at 4°C for 15 mint.
- 5. Transfer the RNA (which is in the supernatant of the tube) without touching the white pallet into a new Eppendorf
- 6. If the sample is small, add 1 μ L (or between 5-10 μ g) of RNase-free glycoblue.
- 7. Add 500µL of isopropanol for 1mL of TRIzol used during cell lysis
- 8. Incubate for 10 mints at RT.
- 9. You can store the the sample at -80°C or continue.

To continue:

- 10. Centrifuge at 10,800 RPM at 4°C for 15 mint. RNA will precipitate into a white pallet
- 11. Remove the supernatant using a pipette
- 12. Wash the pallet with 1mL of 75% ETOH for every 1mL of TRIzol
- 13. The RNA can be stored in 75% ethanol for at least 1 year at –20°C, or at least 1 week at 4°C

To continue:

- 14. Vortex the sample briefly then centrifuge for 5 mints at 8100 RPM at 4°C
- 15. Remove supernatant using a pipette.
- 16. Allow the pallet to airdry for 5-15 mints

IMPORTANT! Do not dry the pellet by vacuum centrifuge. Do not let the RNA pellet dry, to ensure total solubilization of the RNA. Partially dissolved RNA samples have an A230/280 ratio <1.6.

- 17. Resuspend the pallet in 20-50μL of RNAase free water by gently pipetting up & down.
- 18. Incubate in a water bath or heat block set at 55–60°C for 10–15 minutes.
- 19. Dose the RNA using the nanodrop. Blank with RNAase free water and use $2\mu L$ of sample for measurement.
- 20. Record the 260/280 and 230/260 ratios and the RNA quantity in μg .
- 21. Proceed to downstream applications, or store the RNA at -80°C